

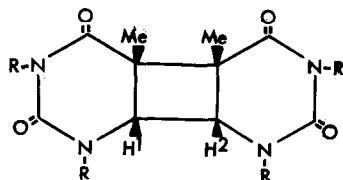
THE PHOTODIMER OF THYMINE: AN NMR STUDY

Ragini Anet

Department of Chemistry, University of California at Los Angeles

(Received 9 August 1965)

The recent communication of Blackburn and Davies¹ prompts us to report our results on the photodimer of thymine (I), obtained² by irradiation of frozen aqueous solutions of thymine. The work described in this paper gives independent confirmation to the head-to-head cis cyclobutane structure for I, predicted³ from the crystal structure of thymine hydrate⁴, and proven by Blackburn and Davies.



I, R=H ; II, R=Me

In the present work, a distinction was made between the four structures⁵ for thymine dimer, by determining the coupling constant $J_{H_1H_2}$ between the two cyclobutane protons, from the naturally abundant C^{13} -H satellite proton spectrum⁶. As I is very sparingly soluble in most solvents, the tetramethyl derivative II, prepared by the methylation of I with dimethylsulphate and alkali⁵, was used. No change in stereochemistry was likely in the conversion of I to II. The methylated derivative II had the further advantage that the $C^{13}H$ satellites of the two N-Methyl groups formed a convenient internal standard for calibration of the cyclobutane

satellite, both for position and intensity. A 25% solution of a highly purified (chromatography, repeated crystallizations) sample of II, m.p. 248°, in deuteriochloroform containing tetramethyl silane was used. The C^{12} -H spectrum was essentially identical with that published⁵. The region, τ 4.34-6.0, was scanned on a Varian A-60 High Resolution NMR spectrometer with a Varian CAT (Computer of Average Transients) attachment, and the sum of 112 scans is shown in Fig. 1.

The cyclobutane satellite was identified as the doublet ($J_{H_1H_2}$, 5 cps) from its position ($J_{C^{13}H}$, 153 ± 2 cps; $J_{N^{13}C-H}$, 139 cps, and 141 cps). The $J_{C^{13}H}$ was expected to be larger than that of cyclobutane⁷ (135 cps) due to the electronegativity of the substituent⁸. Only the low-field cyclobutane satellite could be observed, the one at high field was obscured by the C-CH₃ group satellite.

The low signal-to-noise ratio made a large number of scans necessary to observe the cyclobutane C^{13} -H satellite, thus causing drift and line-broadening. The limited solubility of II precluded the use of more concentrated solutions. The intensity of each component of the doublet (area under the peak) was approximately one-sixth that of the N- $C^{13}H_3$ satellite, and further confirmed its identity as the C^{13} -H satellite of the cyclobutane proton.

The 5 cps coupling ruled out a 1,3 coupling⁹ and showed that I must be a head-to-head dimer. The coupling constant was consistent with a 1,2 cis coupling (5 cps) reported for the dimer of N-methylquinolone¹⁰, but a trans arrangement could not be ruled out.¹¹ However, Weinblum and Johns¹², from a study of the photodimers of thymidine, have concluded that thymine dimer I is a meso, and not a dl compound. Of the two head-to-head structures, only the cis structure is a meso derivative, and hence I must be represented by the head-to-head cis structure I.

The stereochemistry of thymine dimer is similar to that of the photodimer of coumarin¹³. The latter has been shown to arise from the excited singlet state of coumarin by Hammond and co-workers¹⁴. It is tempting to suggest that the photodimer of thymine I formed by U.V. irradiation of DNA in vitro or in vivo¹⁵, arises from the singlet excitation of thymine (cf. Pullman¹⁶). The triplet signal of U.V. irradiated DNA has been shown by Shulman *et al.*¹⁷ to be largely due to thymine anion. The two excited states, singlet thymine, and triplet thymine anion may give rise to different biological effects, namely, repairable thymine dimer formation¹⁸, and a lethal action¹⁹ respectively.

Acknowledgement. I thank Professor P. D. Boyer for his encouragement and support of this work through AEC Contract No. AT-(11-1)-34.

REFERENCES

1. G. M. Blackburn and R. L. H. Davies, *Chemical Communications*, The Chemical Society, 251 (1965).
2. R. Beukers and W. Berends, *Biochim. et Biophys. Acta*, 41, 550 (1960)
3. A. McLaren and D. Shugar in "Photochemistry of Proteins and Nucleic Acids", Pergamon Press, The MacMillan Co., New York (1964).
4. R. Gerdil, *Acta Cryst.*, 14, 333 (1961).
5. D. L. Wulff and G. Fraenkel, *Biochim. et Biophys. Acta*, 51, 332 (1961).
6. A. D. Cohen, N. Sheppard and J. J. Turner, *Proc. Chem. Soc.*, 118 (1958).
7. C. S. Foote, *Tetrahedron Letters*, 9, 579 (1963); J. J. Burke and P. C. Lauterbur, *J. Am. Chem. Soc.*, 86, 1870 (1964).
8. N. Muller, *J. Chem. Phys.*, 36, 359 (1962).
9. P. E. Eaton, *J. Am. Chem. Soc.*, 84, 2344 (1962).
10. O. Buchardt, *Acta Chim. Scand.*, 18, 1389 (1964).
11. V. Georgian, L. Georgian, and A. V. Robertson, *Tetrahedron* 19, 1219 (1963).
12. H. E. Johns, private communication, D. L. Weinblum and H. E. Johns, to be published.

13. R. Anet, *Chem. and Ind.*, 897 (1960); *Can. J. Chem.*, 40, 1249 (1962).
14. G. S. Hammond, C. A. Stout and A. A. Lamola, *J. Am. Chem. Soc.*, 86, 3103 (1964).
15. A. Wacker in "Progress in Nucleic Acid Research", Vol. I, edited by J. N. Davidson and W. E. Cohn, Academic Press, New York (1963).
16. M. J. Mantione and B. Pullman, *Biochim. et Biophys. Acta* 91, 387 (1964).
17. R. O. Rahn, R. K. Shulman and J. W. Longworth, *Proc. Nat. Acad. Sci., Wash.*, 53, 893 (1965).
18. D. Pettijohn and P. Hanawalt, *J. Mol. Biol.* 9, 395 (1964); F. J. Bollum and R. Setlow, *Biochim. et Biophys. Acta*, 68, 599 (1963); R. P. Boyce and P. Howard Flanders, *Proc. Nat. Acad. Sci., Wash.*, 51, 293 (1964).
19. K. C. Smith in *Photophysiology*, Vol. II, edited by A. C. Giese, Academic Press, New York (1964).

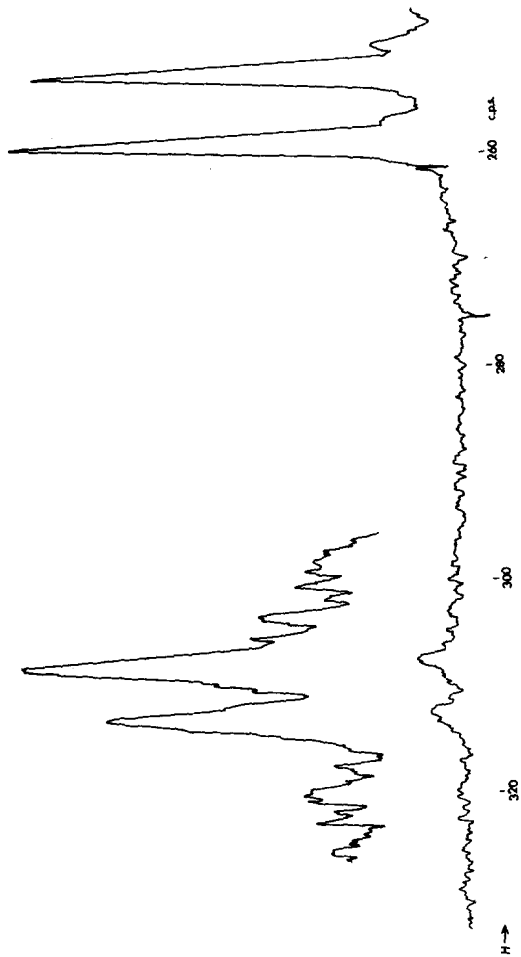


Fig. 1 Low-field C^{13} -H satellite spectrum of the cyclobutane and N-methyl protons of II. C.P.S. are relative to tetramethylsilane. Inset, cyclobutane C^{13} -H satellite.